

Development and Validation of Stability Indicating Method for Estimation of Buparvaquone by Forced Degradation Studies

Shaikh Tanveer Kutubuddin¹, Shelke Sushil Suresh¹, Kakde Rajendra Baliram^{1*}, Lalatsa Aikaterini²

¹Department of Pharmaceutical Sciences, RTMNU, Nagpur, Maharashtra, INDIA.

²School of Pharmacy and Biomedical Sciences, University of Portsmouth, Hampshire, UK.

ABSTRACT

Aim: A lucid, rapid and precise stability-indicating method was developed by using HPLC for the estimation of Buparvaquone in bulk as well as pharmaceutical dosage form by forced degradation studies. **Materials and Methods:** Princeton C₁₈ column (4.6 × 150 mm, 5μ) and mobile phase containing 1% glacial acetic acid and acetonitrile in the proportion of 5:95 v/v was used throughout the study. The flow rate was 0.9 ml/m and the detecting wavelength was kept as 251 nm using the PDA detector. **Results:** The retention time of Buparvaquone was found to be at 8.6 ± 0.5 m. The method developed was validated, as stated in ICH Q2 (R1) guidelines. It was found to be linear within concentration ranging from 2-20 μg/ml having a correlation coefficient 0.999 and other parameters are also under permissible limits. **Conclusion:** Buparvaquone was exposed to different stress conditions like acidic, basic, neutral, thermal, peroxide and also photolytic. Amongst all, the drug was found to be more degraded under basic as well as peroxide conditions.

Key words: Buparvaquone, Forced degradation, Validation, RP-HPLC, Stability.

Key Message: Novel Stability indicating method for the drug buparvaquone has been developed by using HPLC. Forced degradation studies have been carried out and it also reflects the stability of the drug in various stress conditions.

INTRODUCTION

Buparvaquone (BPQ), 2-[4-tert-butylcyclohexyl)methyl]-3-hydroxynaphthalene-1,4-dione (Figure 1), is a second-generation hydroxynaphthoquinone antiprotozoal drug used for the therapy and prophylaxis of all forms of theileriosis.¹ It is also used in the treatment of Cutaneous and Visceral Leishmaniasis.² It was initially developed as an anti-malarial drug³ and is currently the most promising compound for the treatment of theileriosis.⁴

Buparvaquone had displayed high *in-vitro* activity in the case of *L. donovani* infections with ED50 values in the middle of 0.12 and 0.005 μM. But after subcutaneous injection, the *in-vivo* activity of Buparvaquone was low.⁵ Also, the oral bioavailability of BPQ is

low due to low aqueous solubility, which is less than 0.03 μg/ml making the drug highly lipophilic. These problems have led the way to the synthesis of water-soluble phosphate prodrugs of BPQ having water solubility higher than 3.5 μg/ml within the pH range of 3.0-7.4.⁶⁻⁸

Literature Survey discloses that there are specific techniques available for the estimation of BPQ like RP-HPLC-UV method, Spectrofluorimetric method and Bioanalytical method.^{9,10} The current study was designed to develop novel, simple and accurate stability-indicating method^{11,12} for the determination of Buparvaquone in bulk and in pharmaceutical preparation.

Submission Date: 10-10-2019;

Revision Date: 20-04-2020;

Accepted Date: 26-06-2020

DOI: 10.5530/ijper.54.3.131

Correspondence:

Dr. Rajendra B Kakde

Department of Pharmaceuti-

cal Sciences, Rashttrasant

Tukadoji Maharaj Nagpur

University, Nagpur-440033,

Maharashtra, INDIA.

Phone: +91 9822711060

E-mail: drkakde@yahoo.com



www.ijper.org

MATERIALS AND METHODS

Buparvaquone active pharmaceutical ingredient has been obtained as a gift sample from Dr. Lalatsa Lab, University of Portsmouth, Hampshire, UK. Pharmaceutical formulation in the form of tablets (Buparvaquone 100mg) was purchased from the local market manufactured by Vetbiochem India Pvt Ltd, Pune, Maharashtra, India. All the solvents used were of HPLC grade and reagents were of AR grade.

Solvents and Reagents: Acetonitrile, Glacial acetic acid (GAA), water, NaOH, Concentrated HCl, 30% H₂O₂.

Chromatographic conditions

Shimadzu LC-6AD semi-preparative HPLC was used and the mode of elution was isocratic. The mobile phase consisting of acetonitrile and 1% GAA in the proportion of 95:5 v/v was used with flow rate of 0.9 ml/m and the column used was PRINCETON C₁₈ (4.6×150 mm, 5μ). Twenty microliters of the sample have been injected through a loop injector. Detecting wavelength was kept at 251 nm.

Preparation of standard solution

Ten milligram of Buparvaquone was weighed accurately and dissolved in a small quantity of acetonitrile and the final volume was made up to 10ml with the same. It was sonicated for 15 min to get a clear solution (concentration 1000 μg/ml). One ml from the above solution was taken out and diluted up to 100 ml to achieve the final concentration of 10 μg/ml.

Preparation of sample solution

For sample preparation, acetonitrile was used as a solvent for extraction and dilution. Twenty tablets of BPQ were weighed and finely pulverized. An accurately weighed powder of tablet equivalent to 10 mg of Buparvaquone (40.58 mg) was transferred into a 10ml volumetric flask. About 5 ml of acetonitrile was added and the mixture was sonicated for 15 min. The solution was allowed to cool at room temperature and then diluted with acetonitrile until it reaches the mark (stock solution 1000 μg/ml). The above solution is then filtered through Whatman filter paper having grade I. One milliliter of the filtrate was moved to a 100 ml volumetric flask and the volume was made up to the mark using acetonitrile to obtain a concentration of 10 μg/ml.

Method development

For the estimation of Buparvaquone, the pure drug was injected in the HPLC system and run by using the mobile phase. In order to select a mobile phase initially, various

organic solvents in a single and combinations were tried in order to obtain a sharp peak of Buparvaquone with adequate retention. The mobile phase containing plane methanol, plane acetonitrile, a combination of methanol: formic acid, acetonitrile: water, methanol: acetonitrile, formic acid: acetonitrile Combination of phosphate buffer: methanol: acetonitrile at pH range 7 ±0.5 was tried, but satisfactory results were not obtained. Ammonium acetate buffer: methanol and Ammonium acetate buffer: acetonitrile with variable pH range 3.8, 4, 5, 5.8 and different compositions of Ammonium acetate buffer: acetonitrile 10:90, 80:20, 30:70, 40:60, 50:50 was tried. For satisfactory retention time, the flow rate was adjusted at different rates. The mobile phase containing acetonitrile and 1% Glacial acetic acid in the ratio of 95:5 v/v with a flow rate of 0.9 ml/m was found to be competent as it gives a sharp, symmetrical peak with adequate retention.

Validation of method

Validation of the method was carried out as stated in ICH guidelines. The parameters which were evaluated are Linearity, Precision, system suitability, Robustness, Accuracy, Limit of detection (LOD) and Limit of quantitation (LOQ).

System Suitability parameters

The chromatographic conditions were set according to the optimized parameters and the mobile phase was allowed to pass through a stationary phase to get a steady baseline. Six replicate injections of working standard solutions were injected independently and the values were noted.

Linearity

Linearity was studied by injecting the different concentrations of drugs ranging from 2-20 μg/ml. The graph of concentration versus area has been plotted and the correlation coefficient (*r*²) is recorded.

Accuracy

Accuracy of the method was confirmed on the basis of recovery studies carried out by using the standard addition method at three different levels of the labeled claim (80, 100 and 120% w/w). Three replicate injection of each level was made separately and values were recorded.

Precision

The precision of the method has been done by injecting the solution on the same day (Intraday), different day (Interday) and also by using another analyst.

Robustness

The robustness of the proposed method was studied by making small alterations in wavelength and also in the acetonitrile content of the mobile phase. Values were recorded as %RSD.

Limit of detection (LOD) and Limit of quantitation (LOQ)

LOD and LOQ were computed using the formula

$$\text{LOD} = 3.3 \times (\sigma) / s$$

$$\text{LOQ} = 10 \times (\sigma) / s$$

Where,

s means Slope

σ means Standard Deviation (SD)

Forced Degradation Studies

Preparation of Forced Degradation Sample

For the preparation of forced degradation samples, Buparvaquone was subjected to different stress conditions like acidic, alkaline, oxidative, thermal, neutral and photolytic. The stress conditions for each of the degradation types were optimized on trial and error basis to obtain about 5 to 20% degradation.

Acid hydrolysis

Ten mg of Buparvaquone was weighed accurately and solubilized in 10 ml acetonitrile to obtain a concentration of 1000 $\mu\text{g}/\text{ml}$. Then from the above stock solution, 1 ml was diluted up to 10 ml with 2N HCl. This solution is heated under reflux for 18 hr. One ml from the above-heated solution was neutralized with 2N NaOH and again diluted with acetonitrile to achieve a concentration of 10 $\mu\text{g}/\text{ml}$.

Alkaline hydrolysis

Ten mg of BPQ was weighed accurately and then dissolved in 10 ml acetonitrile in order to obtain a concentration of 1000 $\mu\text{g}/\text{ml}$. Then 1 ml from the above-prepared solution was diluted up to 10 ml with 1N NaOH. This solution is heated under reflux for 8 hr.

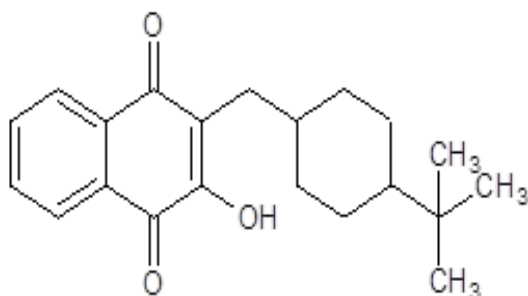


Figure 1: Structure of Buparvaquone.

One ml from the above-heated solution was neutralized with 1N HCl and then again diluted with acetonitrile to achieve the final concentration, which is 10 $\mu\text{g}/\text{ml}$.

Oxidative degradation

Ten mg of BPQ was weighed accurately and then solubilized using 10 ml acetonitrile in order to obtain a concentration of 1000 $\mu\text{g}/\text{ml}$. Then 1 ml from the above solution was diluted up to 10 ml with 30% H_2O_2 . The mixture was placed at room temperature for 24 hr. And to achieve the concentration of 10 $\mu\text{g}/\text{ml}$, one ml from the resultant solution was diluted using acetonitrile.

Neutral hydrolysis

Ten mg of BPQ was weighed accurately and solubilized in 10 ml acetonitrile to obtain a concentration of 1000 $\mu\text{g}/\text{ml}$. Then 1 ml from the above-prepared solution was diluted up to 10 ml with water. The mixture was refluxed on a water bath at 80°C for 24 hr. One ml from the resultant solution was diluted by using acetonitrile to achieve a concentration of 10 $\mu\text{g}/\text{ml}$.

Thermal degradation

An accurately weighed 10 mg of Buparvaquone was kept in an oven at 80°C for 4 hr. After 4 hr, the resultant sample was solubilized in 10 ml acetonitrile to obtain a concentration of 1000 $\mu\text{g}/\text{ml}$. One ml from it was again diluted up to 10 ml with acetonitrile in order to gain the concentration of 100 $\mu\text{g}/\text{ml}$. Then again, 1 ml was taken and diluted up to 10 ml in order to achieve the required concentration of 10 $\mu\text{g}/\text{ml}$.

Photolytic degradation

Ten mg of BPQ was weighed accurately and kept in sunlight for 15 days. After 15 days, the resultant sample was diluted by using 10 ml acetonitrile to obtain a concentration of 1000 $\mu\text{g}/\text{ml}$. One ml from it was then again diluted using acetonitrile up to 10 ml to achieve 100 $\mu\text{g}/\text{ml}$ concentration. Then again, 1 ml was taken and diluted up to 10 ml in order to gain the required concentration of 10 $\mu\text{g}/\text{ml}$.

RESULTS AND DISCUSSION

Method development

Buparvaquone can be easily analyzed by RP-HPLC using the PRINCETON C_{18} column (4.6 \times 150 mm, 5 μ) along with the mobile phase, which is acetonitrile and 1% GAA (95:5 v/v) at a wavelength of 251 nm. The retention time (t_r) of the drug was found at 8.62 min. The flow rate was kept as 0.9 ml/min and the total time of analysis was less than 20 min. The chromatogram obtained for Buparvaquone is shown in Figure 2.

Method Validation System Suitability

From the six replicates of samples injected, it was found that %RSD was under permissible limits, which is less than 2%. The results are listed in Table 1.

Linearity

From the calibration curve (shown in Figure 3), it was found that the method is linear within the concentration ranging from 2-20 µg/ml having a correlation coefficient (R^2) of 0.999. The results are summarized in Table 2.

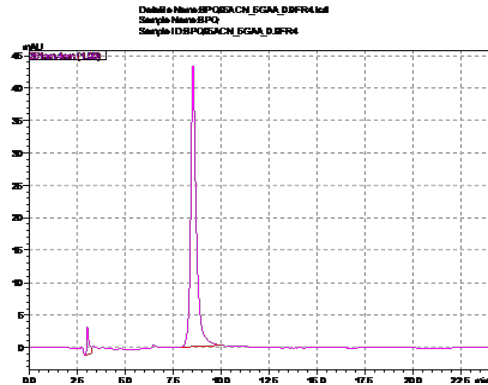


Figure 2: HPLC Chromatogram of Buparvaquone.

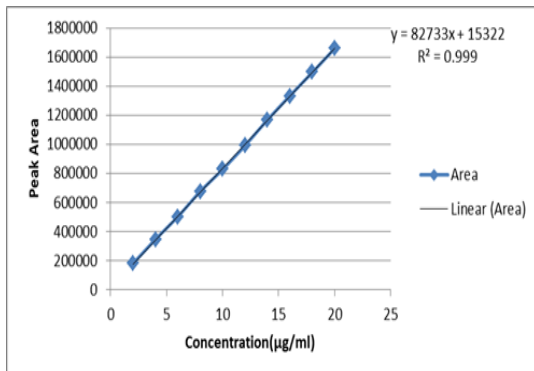


Figure 3: Calibration curve of Buparvaquone.

| Sr. No. | Retention Time (m) | No. of Theoretical Plates | Capacity Factor | Peak Area |
|---------|--------------------|---------------------------|-----------------|-----------|
| 1. | 8.63 | 5368 | 1.351 | 859494 |
| 2. | 8.61 | 5415 | 1.281 | 858245 |
| 3. | 8.64 | 5394 | 1.378 | 858697 |
| 4. | 8.63 | 5376 | 1.328 | 859128 |
| 5. | 8.62 | 5408 | 1.295 | 859492 |
| 6. | 8.63 | 5365 | 1.346 | 858594 |
| Mean | 8.62 | 5387.66 | 1.329 | 858941.7 |
| ± SD | 0.010328 | 21.15341 | 0.036417 | 511.6126 |
| % RSD | 0.119721 | 0.392627 | 1.738431 | 0.0595 |

Accuracy

Recovery studies have been done with the standard addition method and %Recovery near 100% indicates the accuracy of the proposed method. Table 3 shows the results of accuracy.

Precision

Precision studies have been done by estimating the drug concentration and calculating the %RSD on Interday, Intraday and by using different analysts. Interday precision was found to be 0.05%; intraday precision was 0.02% and by using different analysts, the %RSD was found to be 0.09%. The results are shown in Table 4.

Robustness

The proposed method was found to be robust by changing its scanning wavelength and also changing the acetonitrile content in the mobile phase, as shown in Table 5. Results for robustness were shown in Table 6.

LOD and LOQ

LOD and LOQ determination were done with the method based on standard deviation and calibration curve slope. The results were listed in Table 7.

Assay

%Assay was calculated (Table 8) by injecting the sample solution of 10µg/ml in the HPLC system and the area was noted. From that area of sample and area of standard Buparvaquone has been estimated by using the following formula and the summary of validation parameters were shown in Table 9.

$$\% \text{ Assay} = \frac{A_{sam} \times C_{std} \times DF \times \text{Avg.Wt.}}{A_{std} \times \text{Wt.taken} \times LC}$$

Where,

- A_{sam} = Sample area
- A_{std} = Area of standard
- C_{std} = Concentration of standard, µg/ml
- DF = Dilution Factor
- Avg. Wt = Average Weight of tablets
- Wt. taken = Weight of tablet powder taken
- LC = Labeled Claim

| Parameters | Buparvaquone |
|-----------------------------------|----------------------|
| Linear dynamic range (µg/ml) | 2-20 |
| Equation | $Y = 82733X + 15322$ |
| Slope | 82733 |
| Intercept | 15322 |
| Correlation coefficient (R^2) | 0.999 |

| Table 3: Results of Accuracy studies. | | | | | | |
|--|--------------------------|--------------------------|---------|----------|---------------------------|------------|
| Buparvaquone tablets (Avg. Wt. 405.83 mg for 100 mg of Buparvaquone) | | | | | | |
| Level | Wt. of sample taken (mg) | Amount of standard added | Area* | | Total drug estimated (mg) | % Recovery |
| | | | Sample | Standard | | |
| 80% | 40.61 | 8.0 | 1547089 | 859494 | 17.92 | 99.06 |
| 100% | 40.59 | 10.0 | 1718988 | 859494 | 19.96 | 99.64 |
| 120% | 40.60 | 12.0 | 1890886 | 859494 | 21.99 | 99.99 |
| *Mean of 3 readings | | | | Overall | Mean | 99.56 |
| | | | | | ±SD | 0.4691 |
| | | | | | %RSD | 0.4712 |

| Table 4: Results of Precision studies. | | | | |
|--|---------|-------------------|----------|--------------------|
| Sr. No. | Obs. | % Drug estimation | | |
| | | Interday | Intraday | Different Analysts |
| 1. | I | 99.96 | 99.95 | 99.51 |
| 2. | II | 99.85 | 99.99 | 99.46 |
| 3. | III | 99.90 | 99.98 | 99.62 |
| 4. | IV | 99.87 | 99.96 | 99.43 |
| 5. | V | 99.92 | 99.98 | 99.55 |
| 6. | VI | 99.84 | 99.97 | 99.58 |
| | Mean | 99.90 | 99.98 | 99.53 |
| | ±S.D. | 0.0542 | 0.0214 | 0.0953 |
| | %R.S.D. | 0.0543 | 0.0214 | 0.0970 |

| Table 6: Results of Robustness parameters by HPLC. | | | | |
|---|----------------------|--------|--|----------|
| Buparvaquone Tablets (Avg. Wt. 405.83 mg for 100mg of Buparvaquone) | | | | |
| | Wavelength (251±2nm) | | Acetonitrile content in mobile phase [ACN : 1% GAA (95 : 5 v/v)] | |
| | 249nm | 253nm | 93ml ACN | 97ml ACN |
| Mean | 100.51 | 99.89 | 99.86 | 100.15 |
| ±SD | 0.2595 | 0.6644 | 0.412 | 0.2605 |
| %RSD | 0.2501 | 0.6456 | 0.413 | 0.2594 |

| Table 5: Parameters for Robustness study. | | | |
|--|----------|----------|----------|
| Parameter | - Level | Nominal | + Level |
| Change in Scanning Wavelength | 249 nm | 251 nm | 253 nm |
| Change in Acetonitrile content in total mobile phase | 93.00 ml | 95.00 ml | 97.00 ml |

| Table 7: Results of LOD and LOQ studies. | |
|--|------|
| LOD (µg/ml) | 0.47 |
| LOQ(µg/ml) | 1.43 |

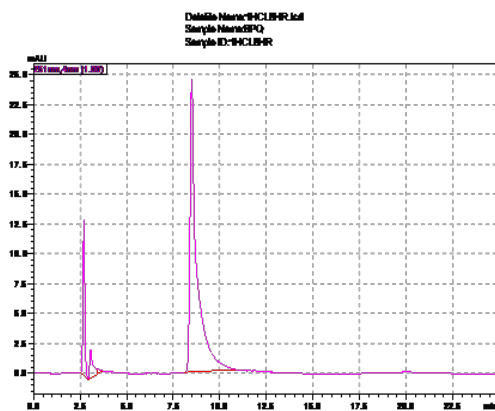


Figure 4: (a) Chromatogram of Buparvaquone after acid hydrolysis.

| Table 8: Assay of marketed formulation. | | | | |
|--|--------------------|----------|--------|---------|
| Buparvaquone tablets (Avg. Wt. 405.83 mg for 100 mg of Buparvaquone) | | | | |
| Sr. No. | Sample weight (mg) | Area | | % Assay |
| | | Standard | Sample | |
| 1 | 40.58 | 859494 | 859218 | 99.97 |
| 2 | 40.54 | 858245 | 858341 | 100.11 |
| 3 | 40.55 | 858697 | 858498 | 100.05 |
| 4 | 40.58 | 859128 | 858998 | 99.99 |
| 5 | 40.55 | 859492 | 859259 | 100.05 |
| Mean | | | | 100.03 |
| SD± | | | | 0.0569 |
| %RSD | | | | 0.0569 |

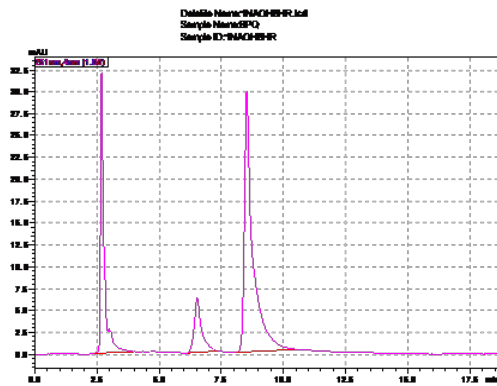


Figure 4: (b) Chromatogram of Buparvaquone after alkaline hydrolysis.

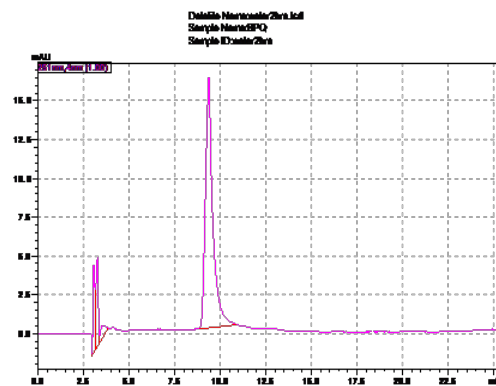


Figure 4: (d) Chromatogram of Buparvaquone after oxidative degradation.

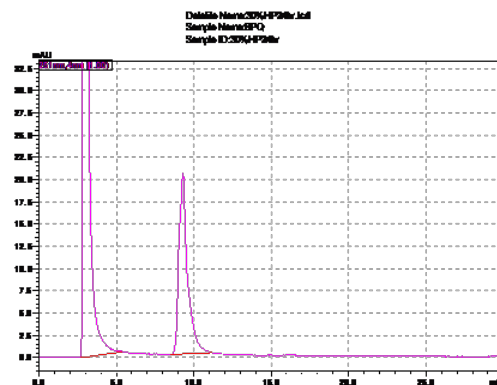


Figure 4: (c) Chromatogram of Buparvaquone after oxidative degradation.

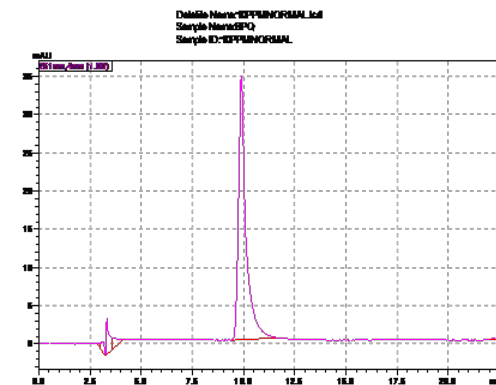


Figure 4: (e) Chromatogram of Buparvaquone after thermal degradation.

Table 9: Summary of Validation Parameters.

| Parameters | Observation |
|------------------------------------|-------------------|
| Linearity and Range | 2-20 µg/ml |
| Regression equation | Y= 82733X + 15322 |
| Intercept | 15322 |
| Slope | 82733 |
| Correlation coefficient (r^2) | 0.999 |
| % Recovery | 99.56% |
| Precision Intraday (%RSD) | 0.02% |
| Precision Interday (%RSD) | 0.05% |
| Precision Different analyst (%RSD) | 0.09% |
| Robustness | Robust |
| LOD | 0.47 µg/ml |
| LOQ | 1.43 µg/ml |
| % Assay | 100.03% |

Forced degradation studies

Forced degradation studies were carried out in order to identify the degradant products. Forced degradation of Buparvaquone was executed under stress conditions of acid, base, oxidation, neutral, thermal and photolytic

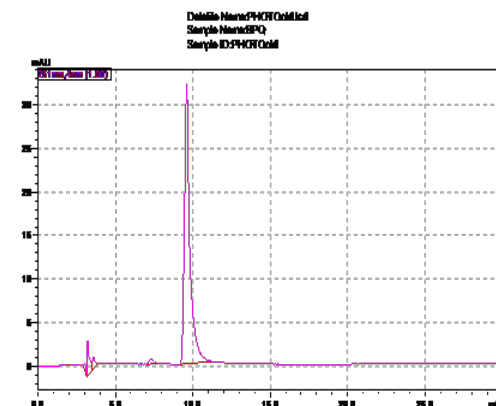


Figure 4: (f) Chromatogram of Buparvaquone after Photolytic degradation.

degradation and the chromatograms are depicted in Figure 4 (a-f). From the results (Table 10), it was clear that the drug was found to be more degraded under alkaline and oxidative conditions.

CONCLUSION

The proposed stability-indicating method was found to be simple, accurate, linear and precise for the estimation of Buparvaquone and the method was

Table 10: Results of forced degradation studies of Buparvaquone.

| | A | B | N | O | T | P |
|--|--------------|-------------|------------------|-----------------------------------|-----------|----------|
| Concentration of stressor | 2N HCl | 1N NaOH | H ₂ O | 30% H ₂ O ₂ | -- | Sunlight |
| Duration of stress conditions | Reflux, 18 h | Reflux, 8 h | Reflux, 24 h | RT, 24 h | 80°C, 4 h | 15 days |
| t _r of Degraded Products(m) | 8.6 | 6.5, 8.6 | 9.2 | 9.4 | 9.8 | 9.2 |
| % of active Buparvaquone | 95.05 | 80.55 | 92.84 | 66.85 | 98.15 | 92.06 |

Key; A: acid; N: neutral; B: base; O: oxidative; T: thermal; P: photolytic

validated according to the guidelines of ICH. The drug gets more degraded under alkaline and oxidative conditions. This method can be utilized for the routine analysis of Buparvaquone in bulk and in pharmaceutical preparation.

ACKNOWLEDGEMENT

The authors would like to thanks Head, Department of Pharmaceutical Sciences, Rashtrasant Tukadoji Maharaj Nagpur University, Nagpur, for providing all the facilities to carry out the research work.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

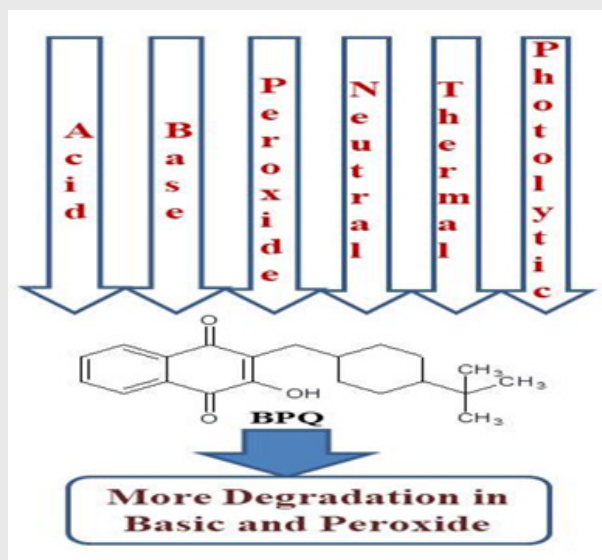
ABBREVIATIONS

HCl: Hydrochloric acid; **NaOH:** Sodium hydroxide; **H₂O₂:** Hydrogen Peroxide; **GAA:** Glacial acetic acid; **RSD:** Relative standard deviation.

REFERENCES

1. Belal FF, Abdel-Razeq SA, Fouad MM, Fouad FA. Densitometric and Spectrometric Determinations of Florfenicol and Buparvaquone. *British Journal of Pharmaceutical Research*. 2014;4(7):806-17.
2. Mantyla A, Garnier T, Rautio J, Nevalainen T, Vepsalainen J, Koskinen A, et al. Synthesis, *in vitro* Evaluation and Antileishmanial Activity of Water-Soluble Prodrugs of Buparvaquone. *J Med Chem*. 2004;47(1):188-95.
3. Hudson A, Randall A, Fry M, Ginger C, Hill B, Latter V, et al. Novel antimalarial hydroxynaphthoquinones with potent broad spectrum antiprotozoal activity. *Parasitology*. 1985;90(01):45-55.
4. Brown C. Control of tropical theileriosis (*Theileria annulata* infection) of cattle. *Parassitologia*. 1990;32(1):23-31.
5. Croft SL, Hogg J, Gutteridge WE, Hudson AT, Randall AW. The Activity of Hydroxynaphthoquinones against *Leishmania donovani*. *J Antimicrob Chem*. 1992;30(6):827-32.
6. Venkatesh G, Ramanathan S, Mansor SM, Nair NK, Sattar MA, Croft SL, et al. Development and validation of RP-HPLC-UV method for simultaneous determination of Buparvaquone, Atenolol, Propranolol, Quinidine and Verapamil: A tool for the standardization of rat *in situ* intestinal permeability studies. *Journal of Pharmaceutical and Biomedical Analysis*. 2007;43(4):1546-51.
7. Venkatesh G, Majid M, Ramanathan S, Mansor SM, Nair NK, Croft SL, et al. Optimization and validation of RP-HPLC-UV method with solid-phase extraction for determination of Buparvaquone in human and rabbit plasma: Application to pharmacokinetic study. *Biomed. Chromatogr*. 2008;22(5):535-41.
8. Venkatesh G, Majid M, Mansor SM, Nair NK, Croft SL, Navaratnam V. *In vitro* and *in vivo* evaluation of self-microemulsifying drug delivery system of Buparvaquone. *Drug Development and Industrial Pharmacy*. 2010;36(6):735-43.
9. Manzoori JL, Amjadi M, Jouyban A, Azar VP, Karami-Bonari AR, Tamizi E. Spectrofluorimetric determination of Buparvaquone in biological fluids, food samples and a pharmaceutical formulation by using Terbium-Deferasirox probe. *Food Chemistry*. 2011;126(4):1845-9.
10. Muraguri GR, Ngumi PN, Wesonga D, Ndungu SG, Wanjohi JM, Bong K, et al. Clinical efficacy and plasma concentrations of two formulations of Buparvaquone in cattle infected with East coast fever (*Theileria parva* infection). *Research in Veterinary Science*. 2006;81(1):119-26.
11. Validation of Analytical Procedures: Text and Methodology, ICH Harmonized Tripartite Guideline, Q2(R1). International Conference on Harmonization, Geneva. 2005;1-13.
12. Stability of New Drug Substances and Products, ICH Harmonized Tripartite Guideline, Q1(R2). International Conference on Harmonization, Geneva. 2005;1-13.

PICTORIAL ABSTRACT



Dr. Rajendra B. Kakde is working as a Professor in Department of Pharmaceutical Sciences, RTMNU, Nagpur India. He also hold the position of Director, RUSA center for bioactives and natural products, RTMNU, Nagpur. He is having 24 years of teaching experience and he has guided 31 M. Pharm students and 8 Ph.D students. He has published 77 research and review articles in various National and International Journals. He recieved many awards and prizes from various organizations.

SUMMARY

The drug buparvaquone gets degraded when it is subjected to various stress conditions. The intensity of degradation is more in basic and oxidative conditions.

About Authors



Mr. Tanveer K. Shaikh is currently working as an Assistant Professor in Raosaheb Patil Danve College of Pharmacy, Badnapur, Dist. Jalna, India. He has completed his M. Pharm in Pharmaceutical Chemistry from Department of Pharmaceutical Sciences, RTMNU, Nagpur, India.



Mr. Sushil S. Shelke has completed his M. Pharm in Pharmaceutical Chemistry from Department of Pharmaceutical Sciences, RTMNU, Nagpur, India.

Cite this article: Shaikh TK, Shelke SS, Kakde RB, Lalatsa A. Development and Validation of Stability Indicating Method for Estimation of Buparvaquone by Forced Degradation Studies. Indian J of Pharmaceutical Education and Research. 2020;54(3):790-7.